

Making Primer and DNA Dilutions

To prepare:

1. Making primer solutions:

Measure the concentration of the primers reverse and forward with the spectrophotometer and calculates like this:

$$V_i = \frac{C_f \times V_f}{C_i}$$

Vi = initial volume
Cf = final concentration
Vf = final volume
Ci = initial concentration

Forward primer:

$$V_i = \frac{10\text{ng}/\mu\text{l} \times 100\mu\text{l}}{866.35\text{ng}/\mu\text{l}}$$

Vi = 1,15 µl primer
in 98,85 µl milliQ water

Reverse primer:

$$V_i = \frac{10\text{ng}/\mu\text{l} \times 100\mu\text{l}}{231.45\text{ng}/\mu\text{l}}$$

Vi = 4,32 µl primer
in 96,68 µl MilliQ water

2. Making DNA samples of 10ng/µl;

Measure the concentration of the stock with the spectrophotometer and dilute the samples to 10ng/µl. Example; the concentration of sample RAM002 is 1969,5 ng/µl. You want a solution with a total volume of 100µl.

$$V_i = \frac{10\text{ng}/\mu\text{l} \times 100\mu\text{l}}{1969.5 \text{ ng}/\mu\text{l}}$$

Vi = 0,51µl Stock
in 99,49µl MilliQ water

Concentrations for PCR reaction

- DNA 10ng/µl
- Buffer 10x
- MgCl₂ 2 mM
- Primer Reverse 50 ng
- Primer Forward 50 ng
- dNTP's 0,2 mM
- Taq polymerase 2,5U

Primer concentration conversion

1 Molar (1M) = (Molecular Weight)g/Liter = 1 mol/Liter

1 μg = 1000ng

1 mg = μg

$\text{Ng}/\mu\text{l}$ = concentration (nmol/ μl) x MW

1 μM = 1 picomoles/ μl = 0.001 nmol/ μl

1 pico = 1000 nano

100 μl = 100 pmol/ μl = 0.1 nmol/ μl

$\text{Ng}/\mu\text{l}$ = 0.1 nmol/ μl x MW